

Notification Date: June 3, 2025 Effective Date: July 10, 2025

# Whole Genome Sequencing for Hereditary Disorders, Varies

Test ID: WGSDX

### **Explanation:**

#### **Current Testing Algorithm**

If a cord blood specimen is received, maternal cell contamination testing will be added and performed at an additional charge.

Skin biopsy or cultured fibroblast specimens: If a skin biopsy is received, fibroblast culture testing will be performed at an additional charge. If viable cells are not obtained, the client will be notified.

New	lesting Algorithm	
None	1	

#### **Current Specimen Required**

**Patient Preparation:** A previous bone marrow transplant from an allogenic donor will interfere with testing. For instructions for testing patients who have received a bone marrow transplant, call 800-533-1710.

Submit only 1 of the following specimens for each family member.

Specimen Type: Whole blood

Container/Tube:

Preferred: Lavender top (EDTA) or yellow top

ACD)

Acceptable: Any anticoagulant Specimen Volume: 3 mL Collection Instructions:

- 1. Invert several times to mix blood.
- 2. Send whole blood specimen in original tube. Do

not aliquot.

Specimen Stability Information: Ambient

(preferred)/Refrigerated Additional Information:

1. If a cord blood specimen is received, MATCC / Maternal Cell Contamination, Molecular Analysis,

#### **New Specimen Required**

**Patient Preparation:** A previous hematopoietic stem cell transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a hematopoietic stem cell transplant.

Submit only 1 of the following specimens for each family member.

Specimen Type: Whole blood

Container/Tube:

**Preferred**: Lavender top (EDTA) or yellow top (ACD)

Acceptable: Green top (Sodium heparin)

Specimen Volume: 3 mL Collection Instructions:

- 1. Invert several times to mix blood.
- 2. Send whole blood specimen in original tube. **Do not**

iquot.

**Specimen Stability Information**: Ambient (preferred) 4 days/Refrigerated 4 days/Frozen 4 days

Additional Information:

1. Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed.

Varies will be performed at an additional charge; maternal blood sample is required.

2. To ensure minimum volume and concentration of DNA is met, the preferred volume of blood must be submitted. Testing may be canceled if DNA requirements are inadequate.

Specimen Type: Skin biopsy

Supplies: Fibroblast Biopsy Transport Media

(T115)

**Container/Tube:** Sterile container with any standard cell culture media (eg, minimal essential media, RPMI 1640). The solution should be supplemented with 1% penicillin and streptomycin.

Specimen Volume: 4-mm punch

Specimen Stability Information: Refrigerated

(preferred)/Ambient

Additional Information: A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks is required to culture fibroblasts before genetic testing can occur.

**Specimen Type: Cultured fibroblast** 

Container/Tube: T-25 flask Specimen Volume: 2 Flasks

**Collection Instructions:** Submit confluent cultured fibroblast cells from a skin biopsy from another laboratory. Cultured cells from a prenatal specimen will not be accepted.

**Specimen Stability Information:** Ambient (preferred)/Refrigerated (<24 hours)

Additional Information: A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks is required to culture fibroblasts before genetic testing can occur.

Specimen Type: Saliva

**Patient Preparation:** Patient should not eat, drink, smoke, or chew gum 30 minutes prior to collection.

Supplies: Saliva Swab Collection Kit (T786)

Specimen Volume: 1 Swab

Collection Instructions: Collect and send

specimen per kit instructions.

**Specimen Stability Information:** Ambient 30

days

Additional Information: Due to lower quantity/quality of DNA yielded from saliva, some aspects of the test may not perform as well as DNA extracted from a whole blood sample. When applicable, specific gene regions that were unable to be interrogated will be noted in the report. Alternatively, additional specimen may be required to complete testing.

Specimen Type: Muscle tissue biopsy Supplies: Muscle Biopsy Kit (T541) 2. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.

Specimen Type: Cord blood

Container/Tube:

Preferred: Lavender top (EDTA) or yellow top (ACD)

Acceptable: Green top (Sodium heparin)

Specimen Volume: 3 mL Collection Instructions:

1. Invert several times to mix blood.

Send cord blood specimen in original tube. Do not aliquot.
 Specimen Stability Information: Ambient (preferred) 4

days/Refrigerated 4 days/Frozen 4 days

#### **Additional Information:**

- 1. Specimens are preferred to be received within 4 days of collection. Extraction will be attempted for specimens received after 4 days, and DNA yield will be evaluated to determine if testing may proceed.
- 2. To ensure minimum volume and concentration of DNA is met, the requested volume must be submitted. Testing may be canceled if DNA requirements are inadequate.
- 3. While a properly collected cord blood sample may not be at risk for maternal cell contamination, unanticipated complications may occur during collection. Therefore, maternal cell contamination studies are recommended to ensure the test results reflect that of the patient tested and are available at an additional charge. Order MATCC / Maternal Cell Contamination, Molecular Analysis, Varies on the maternal specimen.

Specimen Type: Saliva

Patient Preparation: Patient should not eat, drink, smoke, or

chew gum 30 minutes prior to collection. **Supplies**: Saliva Swab Collection Kit (T786)

Specimen Volume: 2 Swabs

Collection Instructions: Collect and send specimen per kit

instructions.

**Specimen Stability Information**: Ambient (preferred) 30

days/Refrigerated 30 days

Additional Information: Saliva specimens are acceptable but not recommended. Due to lower quantity/quality of DNA yielded from saliva, some aspects of the test may not perform as well as DNA extracted from a whole blood sample. When applicable, specific gene regions that were unable to be interrogated will be noted in the report. Alternatively, additional specimen may be required to complete testing.

Specimen Type: Cultured fibroblasts

Source: Skin

Container/Tube: T-25 flask Specimen Volume: 2 Flasks

**Collection Instructions**: Submit confluent cultured fibroblast cells from a skin biopsy. Cultured cells from a prenatal

specimen will not be accepted.

Specimen Stability Information: Ambient (preferred) <24

hours/Refrigerated <24 hours

Additional Information:

1. Specimens are preferred to be received within 24 hours of collection. Culture and extraction will be attempted for

**Collection Instructions:** Prepare and transport specimen per instructions in <u>Muscle Biopsy</u> <u>Specimen Preparation Instructions</u>.

Specimen Volume: 10 to 80 mg

Specimen Stability Information: Frozen

(preferred)/Ambient/Refrigerated

specimens received after 24 hours and will be evaluated to determine if testing may proceed.

2. A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks are required to culture fibroblasts before genetic testing can occur.

Specimen Type: Tissue biopsy Supplies: Hank's Solution (T132)

**Container/Tube**: Sterile container with sterile Hank's balanced salt solution, Ringer's solution, or normal saline

Specimen Volume: 0.5 to 3 cm(3) or larger

Specimen Stability Information: Ambient (preferred) <24

hours/Refrigerated <24 hours

Additional Information:

- 1. Specimens are preferred to be received within 24 hours of collection. Culture and/or extraction will be attempted for specimens received after 24 hours and will be evaluated to determine if testing may proceed.
- 2. A separate culture charge will be assessed under CULFB / Fibroblast Culture for Biochemical or Molecular Testing. An additional 3 to 4 weeks are required to culture fibroblasts before genetic testing can occur.

Specimen Type: Muscle tissue biopsy Supplies: Muscle Biopsy Kit (T541) Specimen Volume: 20 to 80mg

**Collection Instructions:** Prepare and transport specimen per instructions in <u>Muscle Biopsy Specimen Preparation.</u>

Specimen Stability Information: Frozen (preferred) <24

hours/Ambient/Refrigerated <24 hours

**Additional Information:** Specimens are preferred to be received within 24 hours of collection. Extraction will be attempted for specimens received after 24 hours and will be evaluated to determine if testing may proceed.

Specimen Type: Extracted DNA Container/Tube:

Preferred: Screw Cap Micro Tube, 2mL with skirted conical

base

Acceptable: Matrix tube, 1 mL Collection Instructions:

- 1. The preferred volume is at least 100 mcL at a concentration of 75 ng/mcL.
- 2. Include concentration and volume on tube.

Specimen Stability Information: Frozen (preferred) 1

year/Ambient/Refrigerated

Additional Information: DNA must be extracted in a CLIA-certified laboratory or equivalent and must be extracted from a specimen type listed as acceptable for this test (including applicable anticoagulants). Our laboratory has experience with Chemagic, Puregene, Autopure, MagnaPure, and EZ1 extraction platforms and cannot guarantee that all extraction methods are compatible with this test. If testing fails, one repeat will be attempted, and if unsuccessful, the test will be reported as failed and a charge will be applied. If applicable, specific gene regions that were unable to be interrogated due to DNA quality will be noted in the report.

Current Specimen Stability				
Specimen	Temperature	Time	Special Container	
Varies	Ambient (preferred)			
	Frozen			
	Refrigerated			

New Specimen Stability						
Specimen	Temperature	Time	Special Container			
Varies	Varies					

## Questions

Contact Michelle Raths, Laboratory Resource Coordinator at 800-533-1710.